Notizen 159

A Revised Structure for the Phytoalexin Cajanol

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Peroxide oxidation of cajanol ethyl ether afforded the B-ring derived product, 2-methoxy-4-ethoxybenzoic acid. Formation of this compound means that the structure of cajanol, a major phytoalexin from *Cajanus cajan*, must be revised to 5,4'-dihydroxy-7,2'-dimethoxyisoflavanone.

Several isoflavonoid phytoalexins are known to accumulate in the etiolated stems of pigeon pea, Cajanus cajan (Leguminosae; subfamily Papilionoideae; tribe Cajaneae) following inoculation with the fungus, Helminthosporium carbonum [1]. These compounds have been identified [1] as the isoflavones, genistein (5,7,4'-trihydroxy-), 2'-hydroxy-genistein (5,7,2',4'-tetrahydroxy-) and cajanin (5,2',4'-trihydroxy-7-methoxy-) and the isoflavanone, cajanol; the latter phytoalexin was originally formulated as 5,2'-dihydroxy-7,4'-dimethoxyisoflavanone (1).

On chromatograms sprayed with diazotised pnitroaniline, cajanol (M+316; C₁₇H₁₆O₆) immediately gives a bright orange/yellow colouration. In contrast, with Gibbs reagent [2, 3] the expected dark blue indophenol appears slowly, an observation not in accord with an unsubstituted position para to the proposed 2'-hydroxyl function. Thus, the 2'hydroxy isoflavanone, ferreirin (3) (5,7,2'-trihydroxy-4'-methoxyisoflavanone) rapidly affords an intense blue derivative when subjected to the Gibbs test [1]. The abovementioned results suggest that in the B-ring of cajanol, the hydroxyl group may be located at C-4' (and the methoxyl at C-2') rather than C-2' as previously reported [1]. The A-ring assignments (C-5 OH and C-7 OCH₃) - which were made on the basis of established spectral characteristics [4] - and the 2',4'-oxygenation pattern of ring B are not in dispute.

Further studies on the structure of cajanol have recently been facilitated by its isolation in relatively large quantities from the CuCl₂-treated stems (ap-

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prox. 20 mg cajanol/kg fresh wt) and roots (approx. 60 mg/kg fresh wt) of C. cajan (see Experimental). Ethylation of the chromatographically pure isoflavanone and subsequent H₂O₂ oxidation afforded a B-ring derived product indistinguishable by UV, MS and Co-TLC (in 5 solvent systems) from a synthetic specimen of 2-methoxy-4-ethoxybenzoic acid. Formation of this acid allows the B-ring OCH₃ and OH groups of cajanol to be unequivocally located at C-2' and C-4' respectively. Cajanol is thus 5,4'-dihydroxy-7,2'-dimethoxyisoflavanone (2) and not 5,2'-dihydroxy-7,4'-dimethoxyisoflavanone (1) as reported by Ingham [1]. As expected, peroxide oxidation of 5-hydroxy-7,2'-diethoxy-4'-methoxy isoflavanone (7,2'-di-O-ethylferreirin) gave 2-ethoxy-4-methoxybenzoic acid identical (UV, MS and TLC) with an authentic sample. The 2-ethoxy-4-methoxy substituted acid could be readily distinguished from the cajanol-derived isomer by comparative TLC (see Experimental).

1: $R^1 = R^3 = CH_3$; $R^2 = H$

2: R¹=R²=CH₃; R³=H 3: R¹=R²=H; R³=CH₃

4: $R^1 = R^2 = R^3 = H$

5: $R^1 = R^3 = H$; $R^2 = CH_3$

Phytoalexin surveys in the Papilionoideae subfamily of the Leguminosae [5] suggest that cajanol is of exceptionally rare occurrence. Indeed, apart from C. cajan, this isoflavanone has only been obtained from the H. carbonum-inoculated hypocotyls of Florida velvet bean, Mucuna (Stizolobium) deeringianum (tribe Erythrineae) where it co-occurs with various other isoflavonoids including genistein, 2'-hydroxygenistein, dalbergioidin (4) (5,7,2',4'-tetrahydroxyisoflavanone) and a third isoflavanone provisionally identified as 5,7,4'-trihydroxy-2'-methoxyisoflavanone (5) (isoferreirin). In M. deeringianum, the latter compound might represent the immediate biosynthetic precursor of cajanol.

Experimental

Mass and UV spectra were determined as previously described [1]. All chromatographic separations were undertaken using Merck, pre-coated Sigel TLC plates (F 254; layer thickness, 0.25 mm).



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Notizen Notizen

Induction of cajanol

Locally purchased seeds of pigeon pea (Cajanus cajan [L.] Millsp.) were germinated as previously described [1], sown in moist vermiculite and grown (darkness, $24\,^{\circ}\mathrm{C}$) for $15-20\,\mathrm{days}$. The etiolated stems $(200-300\,\mathrm{cm})$ were then cut into short (approx. $10\,\mathrm{cm}$) lengths and placed in clear plastic boxes containing $\mathrm{CuCl_2} \cdot 2\,\mathrm{H_2O}$ $(5\times10^{-3}\,\mathrm{M})$ in 0.05% aqueous Tween 20 to a depth of $3-5\,\mathrm{mm}$. Roots of $C.\,cajan$ were washed with de-ionised $\mathrm{H_2O}$ to remove the vermiculite and then immersed in aqueous $\mathrm{CuCl_2}$ as described above. Stems and roots were incubated [1] for 5 or 6 days prior to extraction of cajanol.

Isolation and purification of cajanol

After incubation, the faintly brown stems (approx. 50 g) were macerated in warm MeOH (60 °C; 300 ml), filtered by suction and the fibrous mat reextracted. The combined filtrates were reduced (in vacuo, 40 °C) to about 15 ml, diluted with deionised H₂O (250 ml) and shaken (×3) with equal volumes of EtOAc. The pooled organic fractions were reduced to dryness and the residue chromatographed in CHCl₃: MeOH (50:1) to afford cajanol as a brown fluorescent band at R_F 0.60. This zone was eluted (MeOH) and re-chromatographed (npentane: Et₂O: HOAc, $75:25:3, \times 3$) to give the pure isoflavanone (1-1.5 mg). Root tissues (approx. 150 g) were treated as described above except that 600 ml MeOH were used for the initial extraction. Final yields of cajanol varied from 8 to 11 mg. Only traces of cajanol were obtained from stems and roots incubated in aqueous Tween 20.

5,4'-dihydroxy-7,2'-dimethoxyisoflavanone (1) (cajanol)

 λ_{max} (nm): MeOH 214 (100%), 229 (94%), 287 (85%), 336 sh (13%); NaOH 215 (100%), 244 (43%), 287 (38%), 356 (24%); NaOAc, Borate and AlCl₃ as lit. [1]; MS as lit. [1] 4'-acetoxy derivative (Py-Ac₂O) (R_F 0.80, CHCl₃). λ_{max} (nm): MeOH 215 (100%), 229 (84%), 288 (78%), 334 sh (10%); AlCl₃ 312, 368; MS (rel. int.) 358 (M⁺; 8), 316 (7), 192 (16), 168 (7), 167 (95), 166 (15), 151 (8), 150 (100), 135 (32), 107 (22).

Ethylation of cajanol

Cajanol (10 mg), dry Me₂CO (5 ml), anhydrous K_2CO_3 (1 g) and diethyl sulphate (15 μ l) were re-

fluxed (approx. 60 °C) for 2 h. After removal of K_2CO_3 (by centrifugation) and Me_2CO (in vacuo, 40 °C), the residue was chromatographed (CHCl₃) to give impure 5-hydroxy-7,2'-dimethoxy-4'-ethoxy-isoflavanone (4'-O-ethylcajanol) at R_F 0.71. Elution and additional TLC in n-pentane: Et_2O : HOAc (75:25:1) afforded about 9 mg of the pure isoflavanone (R_F 0.59).

4'-O-ethylcajanol

 $\lambda_{\rm max}$ (nm): MeOH 212 (100%), 229 (90%), 259 sh (34%), 286 (81%), 338 sh (13%); NaOH 214 (100%), 247 sh (24%), 287 (25%), 356 (6%); AlCl₃ 272, 310, 368; AlCl₃+HCl 270 sh, 307, 366; addition of NaOAc and Borate did not affect the MeOH spectrum; MS (rel. int.) 344 (M⁺; 11), 179 (23), 178 (100), 165 (3), 163 (5), 150 (5), 149 (7), 135 (12), 107 (8). Diazotised p-nitroaniline, yellow/orange; Gibbs reagent, dark blue (colour developing over several min); Fluorescent brown under long wavelength UV light.

H₂O₂ oxidation of 4'-O-ethylcajanol

The above compound (2 mg), aqueous KOH $(10\%; 3 \text{ ml}), \text{ EtOH } (0.5 \text{ ml}) \text{ and } \text{H}_2\text{O}_2 (30\%;$ 0.2 ml added in $20 \mu l$ portions over 1 h) were stirred $(50\,^{\circ}\text{C}\pm2\,^{\circ}\text{C})$ for 90 min. The mixture was then acidified (2 N HCl; pH 3), extracted (\times 3) with equal volumes EtOAc and the organic fractions pooled and reduced to dryness. TLC (CHCl₃: MeOH, 50:1) of the residue gave a broad, fluorescencequenching band $(R_F 0.44)$ opposite the marker of 2-methoxy-4-ethoxybenzoic acid authentic authentic 2-ethoxy-4-methoxybenzoic acid, R_F 0.56). Traces of unchanged 4'-O-ethylcajanol were detected (diazotised p-nitroaniline) at R_F 0.90. Further TLC purification (n-pentane: Et₂O: HOAc, 75:25:3) of the R_F 0.44 fraction gave 2-methoxy-4-ethoxybenzoic acid (R_F 0.32; cf. 2-ethoxy-4-methoxybenzoic acid, R_F 0.46) indistinguishable (UV, MS, TLC) from synthetic material.

2-methoxy-4-ethoxybenzoic acid

 $\lambda_{\rm max}$ (nm) EtOH: 214 (100%), 254 (81%), 289 (46%); MS (rel. int.) 196 (M+; 70), 179 (5), 167 (7), 151 (65), 139 (37), 138 (11), 123 (8), 122 (25), 121 (100), 108 (10). The acid derived from 4'-O-ethylcajanol co-chromatographed with authentic material in the following solvents: (a) CHCl₃: CCl₄ (3:1), R_F 0.11 (b) C_6H_6 : MeOH (9:1), R_F 0.27

Notizen 161

and (c) Et₂O: n-hexane (3:1), R_F 0.20. Corresponding R_F values for 2-ethoxy-4-methoxybenzoic acid were 0.23, 0.43 and 0.36.

Ethylation of ferreirin

Ferreirin (12 mg) was ethylated using the procedure described for cajanol. TLC purification (CHCl₃: CCl₄, 3:1) of the reaction products gave about 10 mg of 5-hydroxy-7,2'-diethoxy-4'-methoxy-isoflavanone (7,2'-di-O-ethylferreirin) (R_F 0.25) together with small quantities (about 1.5 mg) of 5,2'-dihydroxy-7-ethoxy-4'-methoxyisoflavanone (7-O-ethylferreirin) (R_F 0.06).

7-O-ethylferreirin

 $\lambda_{\rm max} \ ({\rm nm}) : \ MeOH \ 215 \ (100\%), \ 229 \ {\rm sh} \ (95\%), \ 287 \ (90\%), \ 340 \ {\rm sh} \ (15\%) ; \ NaOH \ 218 \ (100\%), \ 245 \ {\rm sh} \ (53\%), \ 289 \ (61\%), \ 352 \ (34\%) ; \ AlCl_3 \ 274 \ {\rm sh}, \ 311, \ 368; \ AlCl_3 + HCl \ 274 \ {\rm sh}, \ 309, \ 368; \ {\rm addition} \ {\rm of} \ NaOAc \ {\rm and} \ Borate \ did \ not \ affect \ the \ MeOH \ {\rm spectrum}; \ MS \ ({\rm rel.\,int.}) \ 330 \ (M^+; \ 24), \ 182 \ (8), \ 181 \ (100), \ 178 \ (27), \ 163 \ (10), \ 153 \ (16), \ 151 \ (7), \ 150 \ (47), \ 149 \ (11), \ 148 \ (5), \ 124 \ (5), \ 121 \ (5). \ Diazotised \ p-nitroaniline, \ bright \ yellow; \ Gibbs \ reagent, \ deep \ blue \ (colour \ developing \ rapidly); \ Fluorescent \ orange/brown \ under \ long \ wavelength \ UV \ light.$

7,2'-di-O-ethylferreirin

 λ_{max} (nm): MeOH 214 (100%), 228 (93%), 287 (87%), 338 sh (16%); NaOH 216 (100%), 246 sh

[1] J. L. Ingham, Z. Naturforsch. 31 c, 504 (1976).

 $(52\%),\,287\,\,(68\%),\,357\,\,(23\%)\,;\, {\rm AlCl_3}\,\,274\,\,{\rm sh},\,312,\,370\,;\,\,{\rm AlCl_3}\,+\,{\rm HCl}\,\,274\,\,{\rm sh},\,309,\,\,370\,;\,\,{\rm addition}\,\,{\rm of}\,\,{\rm NaOAc}\,\,{\rm and}\,\,{\rm Borate}\,\,{\rm did}\,\,{\rm not}\,\,{\rm affect}\,\,{\rm the}\,\,{\rm MeOH}\,\,{\rm spectrum}\,;\,\,{\rm MS}\,\,({\rm rel.\,int.})\,\,358\,\,({\rm M}^+;\,15),\,179\,\,(13),\,178\,\,(100),\,177\,\,(5),\,165\,\,(6),\,164\,\,(5),\,163\,\,(56),\,150\,\,(15),\,149\,\,(32),\,135\,\,(5),\,121\,\,(5).\,\,{\rm Diazotised}\,\,p{\rm nitroaniline},\,{\rm pale}\,\,{\rm yellow}\,;\,\,{\rm Gibbs}\,\,{\rm reagent},\,\,{\rm weak}\,\,{\rm blue}\,;\,\,{\rm Fluorescent}\,\,\,{\rm orange/brown}\,\,\,{\rm under}\,\,\,{\rm long}\,\,\,{\rm wavelength}\,\,\,{\rm UV}\,\,{\rm light}.$

H₂O₂ oxidation of 7,2'-di-O-ethylferreirin

The above isoflavanone (5 mg) was treated with $\mathrm{H_2O_2}$ as described for 4'-O-ethylcajanol. TLC purification of the product (see oxidation of 4'-O-ethylcajanol for solvents and R_F values) gave 2-ethoxy-4-methoxybenzoic acid indistinguishable (UV, MS, TLC) from an authentic sample.

2-ethoxy-4-methoxybenzoic acid

 $\begin{array}{l} \lambda_{\rm max} \ ({\rm nm}) \colon EtOH \ 215 \ (100\%), \ 254 \ (78\%), \ 289 \\ (35\%) \colon MS \ \ ({\rm rel. \, int.}) \ \ 196 \ \ (M^+; \ 33), \ 163 \ \ (20), \\ 152 \ \ (7), \ 151 \ \ (19), \ 150 \ \ (100), \ 135 \ \ (6), \ 122 \ \ (48), \\ 107 \ \ (23). \ For \ comparative \ TLC \ data \ see \ 2-methoxy-4-ethoxybenzoic \ acid. \end{array}$

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[5] J. L. Ingham, Unpublished data.

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^[4] T. J. Mabry, K. R. Markham, and M. B. Thomas, The Systematic Identification of Flavonoids, Springer, New York 1970.